

Package ‘bambu’

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Type Package

Title Reference-guided isoform reconstruction and quantification for long read RNA-Seq data

Version 0.2.0

Description Multi-sample transcript discovery and quantification using long read RNA-Seq data.

License GPL-3

Encoding UTF-8

LazyData true

Depends R(>= 4.0.0),
data.table(>= 1.1.8),
dplyr,
SummarizedExperiment(>= 1.1.6),
GenomicRanges,
BiocManager,
GenomicFeatures,
ggplot2

Suggests knitr,
rmarkdown,
fs,
testthat,
ComplexHeatmap,
circlize,
ggbio,
RColorBrewer,
gridExtra,
BSgenome.Hsapiens.NCBI.GRCh38,
TxDb.Hsapiens.UCSC.hg38.knownGene,
AnnotationDbi,
BSgenome,
BiocGenerics,
BiocParallel,
Biostrings

Enhances parallel

SystemRequirements

biocViews FeatureExtraction,
GeneExpression,
GenomeAnnotation,
ImmunoOncology,
Normalization,
RNASeq,
Regression,
Sequencing,
Software,
Transcription,
Transcriptomics

bugReports <https://github.com/GoekeLab/bambu/issues>

URL <https://github.com/GoekeLab/bambu>

RoxygenNote 7.1.0

LinkingTo Rcpp,
RcppArmadillo,
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Imports S4Vectors(>= 0.22.1),
IRanges,
GenomicAlignments,
glmnet,
Rsamtools,
Rcpp,
RcppArmadillo,
RcppProgress,
progress

VignetteBuilder knitr,
rmarkdown

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bambu

*long read isoform reconstruction and quantification***Description**

This function takes bam file of genomic alignments and performs isoform reconstruction and gene and transcript expression quantification. It also allows saving of read class files of alignments, extending provided annotations, and quantification based on extended annotations. When multiple samples are provided, extended annotations will be combined across samples to allow comparison.

Usage

```
bambu(
  reads = NULL,
  readClass.file = NULL,
  readClass.outputDir = NULL,
  annotations = NULL,
  genomeSequence = NULL,
  stranded = FALSE,
  ncore = 1,
  yieldSize = NULL,
  isoreParameters = NULL,
  emParameters = NULL,
  extendAnnotations = TRUE,
  verbose = FALSE
)
```

Arguments

- | | |
|---------------------|---|
| reads | A string or a vector of strings specifying the paths of bam files for genomic alignments, or a BamFile object or a BamFileList object (see Rsamtools). |
| readClass.file | A string or a vector of strings specifying the read class files that are saved during previous run of bambu . |
| readClass.outputDir | A string variable specifying the path to where read class files will be saved. |
| annotations | A TxDb object or A GRangesList object obtained by prepareAnnotations or prepareAnnotationsFromGTF . |
| genomeSequence | A fasta file or a BSGenome object. |
| stranded | A boolean for strandedness, defaults to FALSE. |
| ncore | specifying number of cores used when parallel processing is used, defaults to 1. |
| yieldSize | see Rsamtools . |
| isoreParameters | A list of controlling parameters for isoform reconstruction process: <ul style="list-style-type: none"> • prefix specifying prefix for new gene Ids (genePrefix.number), defaults to empty |

- `remove.subsetTx` indicating whether filter to remove read classes which are a subset of known transcripts(), defaults to TRUE
 - `min.readCount` specifying minimum read count to consider a read class valid in a sample, defaults to 2
 - `min.readFractionByGene` specifying minimum relative read count per gene, highly expressed genes will have many high read count low relative abundance transcripts that can be filtered, defaults to 0.05
 - `min.sampleNumber` specifying minimum sample number with minimum read count, defaults to 1
 - `min.exonDistance` specifying minimum distance to known transcript to be considered valid as new, defaults to 35
 - `min.exonOverlap` specifying minimum number of bases shared with annotation to be assigned to the same gene id, defaults 10 base pairs
- `emParameters` A list of controlling parameters for quantification algorithm estimation process:
- `maxiter` specifying maximum number of run iterations, defaults to 10000.
 - `bias` specifying whether to correct for bias, defaults to FALSE.
 - `conv` specifying the convergence threshold control, defaults to 0.0001.
- `extendAnnotations` A logical variable indicating whether annotations are to be extended for quantification.
- `verbose` A logical variable indicating whether processing messages will be printed.

Details

Main function

Value

A list of two SummarizedExperiment object for transcript expression and gene expression.

Examples

```
## =====
## Minimum read support 5
## Increase EM convergence threshold to 10^(-6)
test.bam <- system.file("extdata",
  "SGNex_A549_directRNA_replicate5_run1_chr9_1_1000000.bam",
  package = "bambu")
gr <- readRDS(system.file("extdata",
  "annotationGranges_txdbGrch38_91_chr9_1_1000000.rds",
  package = "bambu"))
fa.file <- system.file("extdata",
  "Homo_sapiens.GRCh38.dna_sm.primary_assembly_chr9_1_1000000.fa",
  package = "bambu")
se = bambu(reads = test.bam, annotations = gr,
  genomeSequence = fa.file, extendAnnotations = FALSE)
```

plot.bambu

*plot.bambu***Description**

plotSEOutput

Usage

```
## S3 method for class 'bambu'
plot(
  se,
  group.variable = NULL,
  type = c("annotation", "pca", "heatmap"),
  gene_id = NULL,
  transcript_id = NULL
)
```

Arguments

<code>se</code>	An summarized experiment object obtained from bambu or transcriptToGeneExpression .
<code>group.variable</code>	Variable for grouping in plot, has to be provided if choosing to plot PCA.
<code>type</code>	plot type variable, a values of annotation for a single gene with heatmap for isoform expressions, pca, or heatmap, see details.
<code>gene_id</code>	specifying the <code>gene_id</code> for plotting gene annotation, either <code>gene_id</code> or <code>transcript_id</code> has to be provided when <code>type = "annotation"</code> .
<code>transcript_id</code>	specifying the <code>transcript_id</code> for plotting transcript annotation, either <code>gene_id</code> or <code>transcript_id</code> has to be provided when <code>type = "annotation"</code>

Details

[type](#) indicates the type of plots to be plotted. There are two types of plots can be chosen, PCA or heatmap.

Value

A heatmap plot for all samples

Examples

```
se <- readRDS(system.file("extdata",
  "seOutputCombined_SGNex_A549_directRNA_replicate5_run1_chr9_1_1000000.rds",
  package = "bambu"))
colnames(se) <- colData(se)$name <- c("sample1", "sample2")
assays(se)$CPM[,2] <- pmax(0, rnorm(length(assays(se)$CPM[,2]),
assays(se)$CPM[,2], 10))
plot.bambu(se, type = "heatmap")
```

prepareAnnotations	<i>prepare annotations from txdb object</i>
--------------------	---

Description

Function to prepare tables and genomic ranges for transcript reconstruction using a txdb object

Usage

```
prepareAnnotations(txdb)
```

Arguments

txdb a [TxDb](#) object

Value

A [GRangesList](#) object

Examples

```
library(TxDb.Hsapiens.UCSC.hg38.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg38.knownGene
prepareAnnotations(txdb)
```

```
prepareAnnotationsFromGTF
```

Prepare annotation granges object from GTF file into a GRangesList object

Description

Prepare annotation granges object from GTF file

Usage

```
prepareAnnotationsFromGTF(file)
```

Arguments

file a GTF file

Details

Unlike `readFromGTF`, this function finds out the equivalence classes between the transcripts, with `mcols` data having three columns:

- TXNAME specifying prefix for new gene Ids (`genePrefix.number`), defaults to empty
- GENEID indicating whether filter to remove read classes which are a subset of known transcripts(), defaults to TRUE
- eqClass specifying minimum read count to consider a read class valid in a sample, defaults to 2

Value

A `GRangesList` object

Examples

```
gtf.file <- system.file("extdata",
  "Homo_sapiens.GRCh38.91_chr9_1_1000000.gtf",
  package = "bambu")
gr <- prepareAnnotationsFromGTF(gtf.file)
```

<code>readFromGTF</code>	<i>convert a GTF file into a GRangesList</i>
--------------------------	--

Description

Outputs `GRangesList` object from reading a GTF file

Usage

```
readFromGTF(file)
```

Arguments

`file` a .gtf file

Value

`grlist` a `GRangesList` object, with two columns

- TXNAME specifying prefix for new gene Ids (`genePrefix.number`), defaults to empty
- GENEID indicating whether filter to remove read classes which are a subset of known transcripts(), defaults to TRUE

Examples

```
gtf.file <- system.file("extdata",
  "Homo_sapiens.GRCh38.91_chr9_1_1000000.gtf",
  package = "bambu")
readFromGTF(gtf.file)
```

transcriptToGeneExpression	<i>transcript to gene expression</i>
----------------------------	--------------------------------------

Description

Reduce transcript expression to gene expression

Usage

```
transcriptToGeneExpression(se)
```

Arguments

se a SummarizedExperiment object from [bambu](#)

Value

A SummarizedExperiment object

Examples

```
se <- readRDS(system.file("extdata",  
  "seOutput_SGNex_A549_directRNA_replicate5_run1_chr9_1_1000000.rds",  
  package = "bambu"))  
transcriptToGeneExpression(se)
```

writeBambuOutput	<i>Write bambu results to GTF and transcript/gene-count files</i>
------------------	---

Description

Outputs a GTF file, transcript-count file, and gene-count file from bambu

Usage

```
writeBambuOutput(se, path)
```

Arguments

se a [SummarizedExperiment](#) object from [bambu](#)
path the destination of the output files (gtf, transcript counts, and gene counts)

Value

The function will generate three files, a .gtf file for the annotations, two .txt files for transcript and gene counts respectively.

Examples

```
se <- readRDS(system.file("extdata",
  "seOutput_SGNex_A549_directRNA_replicate5_run1_chr9_1_1000000.rds",
  package = "bambu"))
path <- tempdir()
writeBambuOutput(se, path)
```

writeToGTF	<i>write GRangeslist into GTF file</i>
------------	--

Description

Write annotation GRangesList into a GTF file

Usage

```
writeToGTF(annotation, file, geneIDs = NULL)
```

Arguments

annotation	a GRangesList object
file	the output gtf file name
geneIDs	an optional dataframe of geneIDs (column 2) with the corresponding transcriptIDs (column 1)

Value

gtf a GTF dataframe

Examples

```
outputGtfFile <- tempfile()
gr <- readRDS(system.file("extdata",
  "annotationGranges_txdbGrch38_91_chr9_1_1000000.rds",
  package = "bambu"))
writeToGTF(gr, outputGtfFile)
```

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